JC18 Rec'd PCT/PTO 09 JUL 2001

FORM PTO 1390 (REV 5-93) US DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY DOCKET NUMBER
2001_0971A

TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. §371

U.S. APPLICATION NO. (if known, see 37 CFR 1.5)
09/869917
NEW

International Application No.	International Filing Date	Priority Date Claimed
PCT/JP00/03550	June 1, 2000	December 14, 1999

Title of Invention
IMMUNOASSAY OF PIVKA-II

Applicant(s) For DO/EO/US
Motohito KANASHIMA, Tomohide ASAI, Hiroki TAKAHASHI, Yoshiyuki ASAI

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. [X] This is a **FIRST** submission of items concerning a filing under 35 U.S.C. §371.

2. [] This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. §371.

3. [] This express request to begin national examination procedures (35 U.S.C. §371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. §371(b) and PCT Articles 22 and 39(1).

4. [] A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.

5. [X] A copy of the International Application as filed (35 U.S.C. §371(c)(2))
   a. [] is transmitted herewith (required only if not transmitted by the International Bureau).
   b. [X] has been transmitted by the International Bureau.
   c. [] is not required, as the application was filed in the United States Receiving Office (RO/US)

6. [X] A translation of the International Application into English (35 U.S.C. §371(c)(2)). **ATTACHMENT A**

7. [] Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. §371(c)(3)).
   a. [] are transmitted herewith (required only if not transmitted by the International Bureau).
   b. [] have been transmitted by the International Bureau.
   c. [] have not been made; however, the time limit for making such amendments has NOT expired.
   d. [] have not been made and will not be made.

8. [] A translation of the amendments to the claims under PCT Article 19.

9. [X] An oath or declaration of the inventor(s) (35 U.S.C. §371(c)(4)). **ATTACHMENT B**

10. [] A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. §371(c)(5)).

Items 11. to 14. below concern other document(s) or information included:

11. [X] An Information Disclosure Statement under 37 CFR 1.97 and 1.98. **ATTACHMENT C**

12. [X] An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.

    ATTACHMENT D

13. [X] A FIRST preliminary amendment. **ATTACHMENT E**

    [] A SECOND or SUBSEQUENT preliminary amendment.

14. [] Other items or information:

THE COMMISSIONER IS AUTHORIZED TO CHARGE ANY DEFICIENCY IN THE FEE FOR THIS PAPER TO DEPOSIT ACCOUNT NO. 23-0975.

JC18 Rec'd PCT/PTO 09 JUL 2001

U.S. APPLICATION NO. (If known, see 37 CFR 1.5) NEW 09/869917	INTERNATIONAL APPLICATION NO. PCT/JP00/03550	ATTORNEY'S DOCKET NO. 2001_0971A

15. [X] The following fees are submitted

	CALCULATIONS	PTO USE ONLY
BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)):		
Neither international preliminary examination fee nor international search fee paid to USPTO and International Search Report not prepared by the EPO or JPO $1000.00		
International Search Report has been prepared by the EPO or JPO ..... $ 860.00		
International preliminary examination fee not paid ot USPTO but international search paid to USPTO ..... $ 710.00		
International preliminary examination fee paid to USPTO but claims did not satisfy provisions of PCT Article 33(1)-(4) ..... $ 690.00		
International preliminary examination fee paid ot USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) ..... $ 100.00		
ENTER APPROPRIATE BASIC FEE AMOUNT =	$860.00	
Surcharge of $130.00 for furnishing the oath or declaration later than [] 20 [] 30 months from the earliest claimed priority date (37 CFR 1.492(e)).	$	

Claims	Number Filed	Number Extra	Rate		
Total Claims	-20 =		X $18.00	$	
Independent Claims	- 3 =		X $80.00	$	
Multiple dependent claim(s) (if applicable)			+ $270.00	$	
TOTAL OF ABOVE CALCULATIONS =				$860.00	
[] Small Entity Status is hereby asserted. Above fees are reduced by 1/2.				$	
SUBTOTAL =				$860.00	
Processing fee of $130.00 for furnishing the English translation later than [] 20 [] 30 months from the earliest claimed priority date (37 CFR 1.492(f)). +				$	
TOTAL NATIONAL FEE =				$860.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). $40 per property +				$40.00	
TOTAL FEES ENCLOSED =				$900.00	
				Amount to be refunded	$
				Amount to be charged	$

a. [X] A check in the amount of $900.00 to cover the above fees is enclosed. A duplicate copy of this form is enclosed.

b. [] Please charge my Deposit Account No. 23-0975 in the amount of $_____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. [] The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 23-0975.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

19. CORRESPONDENCE ADDRESS

000513

PATENT TRADEMARK OFFICE

By: Warren Cheek

Warren M. Cheek, Jr.
Registration No. 33,367

WENDEROTH, LIND & PONACK, L.L.P.
2033 "K" Street, N.W., Suite 800
Washington, D.C. 20006-1021
Phone:(202) 721-8200
Fax:(202) 721-8250

July 9, 2001

[CHECK NO. 45353]

[2001_0971A]

09/869917
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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of :

Motohito KANASHIMA et al. : **Attn: BOX PCT**

Serial No. NEW : **Docket No. 2001_0971A**

Filed July 9, 2001 :

IMMUNOASSAY OF PIVKA-II
[Corresponding to PCT/JP00/03550
Filed June 1, 2000]

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents,
Washington, DC 20231

Sir:

Prior to calculating the filing fee, please amend the above-identified application as follows:

IN THE SPECIFICATION

Page 1, immediately after the title, please insert:

This application is a 371 of PCT/JP00/03550 filed June 1, 2000.

IN THE CLAIMS

Please amend the claims as follows:

3. (Amended) The immunoassay according to claim 1, wherein an anti-fibrinogen antibody and/or an anti-fibrin antibody are used as the antibody reacting with human fibrin-like related substances.

ATTACHMENT E

Please add the following new claim:

4. The immunoassay according to claim 1, wherein an anti-fibrinogen antibody and/or an anti-fibrin antibody are used as the antibody reacting with human fibrin-like related substances.

REMARKS

The foregoing amendments amend the specification to reflect the 371 status. In addition, the multiple dependency of the claims has been removed to reduce the PTO filing fee.

Attached hereto is a marked-up version of the changes made to the specification and claims by the current amendment. The attached pages are captioned "**Version with markings to show changes made**".

Favorable action on the merits is solicited.

Respectfully submitted,

Motohito KANASHIMA et al.

By _____
Warren M. Cheek, Jr.
Registration No. 33,367
Attorney for Applicants

WMC/dlk
Washington, D.C. 20006-1021
Telephone (202) 721-8200
Facsimile (202) 721-8250
July 9, 2001

Version with Markings to Show Changes Made

DESCRIPTION

IMMUNOASSAY OF PIVKA-II

This application is a 371 of PCT/JP00/03550 filed June 1, 2000.

Technical Field:

The present invention relates to an immunoassay through an antigen-antibody reaction for specifically and highly sensitively measuring PIVKA-II (Protein Induced by Vitamin K Absence or Antagonist-II) in serum or plasma by adding to reagents thrombin and/or an antibody reacting with human fibrin-like related substances.

Background Art:

Along with AFP(α -fetoprotein), PIVKA-II (Protein Induced by Vitamin K Absence or Antagonist-II) is measured widely in clinical examination laboratories as a hepatic cell tumor detecting marker which specifically increases in hepatic cell cancer patients. Generally, magnetic beads, glass beads, plastic plates, latexes or the like on which PIVKA-II specific monoclonal or polyclonal antibodies are adsorbed are subjected to a first reaction with serum or plasma, then after being washed for B/F separation, a second reaction where human prothrombin specific polyclonal or monoclonal antibodies labeled with an enzyme, a fluorescent material, a radioisotope, an Ru complex or the like are added is carried out, and further after being washed for B/F

Version with Markings to Show Changes Made

CLAIMS

1. An immunoassay of PIVKA-II in serum or plasma, comprising the steps of:

adding to reagents thrombin and/or an antibody reacting with human fibrin-like related substances, and

measuring PIVKA-II in serum or plasma.

2. The immunoassay according to claim 1, wherein a thrombin-containing animal serum and/or purified thrombin are used as the thrombin.

3. (Amended) The immunoassay according to claim 1~~ or 2~~, wherein an anti-fibrinogen antibody and/or an anti-fibrin antibody are used as the antibody reacting with human fibrin-like related substances.

09/869917
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IMMUNOASSAY OF PIVKA-II

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separation, absorbance or luminescence of the enzyme, the fluorescent material, the radioisotope or the Ru bound to an immune complex formed through the antigen-antibody reaction being measured to determine PIVKA-II in the serum or the plasma.

Heretofore, PIVKA-II has been measured by an enzyme immunoassay (EIA), but the EIA is poor in sensitivity with a low positive rate for a relatively small hepatoma, so that an electrochemiluminescence immunoassay (ECLIA) where an antigen or an antibody is labeled with an Ru complex has been recently developed for further highly sensitive measurement. The application of the electrochemiluminescence immunoassay led successfully to higher sensitivity in the PIVKA-II measurement. To realize higher sensitivity not only in the ECLIA but also in an enzyme immunoassay, a chemiluminescence assay, a radioisotope assay, latex turbidimetry or the like, the influence of an unspecific reaction in a sample should be taken into consideration.

In the process of studies for eliminating the influence of the unspecific reaction in a sample in the PIVKA-II measurement, it has been found that sensitivity and specificity of the measurement could be improved by adding to reagents thrombin and/or an antibody reacting highly sensitively with human fibrin-like related substances. As the substances attributable to such an unspecific reaction in the sample, attention was directed first to fibrin or its related substances in the sample and second to

thrombin bound to fibrin or its related substances. In particular, when a polyclonal antibody is used as an anti-human prothrombin antibody for a second antibody or a labeled antibody, it may be subject to the interference of these unspecific reaction substances to cause positive errors in measurement of PIVKA-II. It is reported that the protein structure of prothrombin is composed of an F_1 fragment, an F_2 fragment and thrombin. The labeled antibody used for measurement of PIVKA-II may be not only an anti-prothrombin antibody but also an anti-F_1 antibody, an anti-F_2 antibody, or an anti-(F_1 + F_2) antibody. However, in consideration of the purity of these antibodies or the similarity of thrombin to the antigen, these antibodies may also react with bound or free thrombin in a sample. Further, in measurement of PIVKA-II, fibrin or its insoluble related substances in a sample are physically adsorbed onto carriers such as magnetic beads, glass beads, latexes, plastic plates or the like to give rise to the phenomenon of positive errors in the measurement.

In order to prevent the interference attributable to the fibrin-like related substances and the interference attributable to thrombin, antibodies reacting with the human fibrin-like related substances, for example, anti-fibrinogen or anti-fibrin antibodies and/or thrombin are added to reagents, thereby succeeding in accurately measuring a very small amount of PIVKA-II while effectively inhibiting the nonspecific reaction, leading to the achievement of the present invention.

An object of the present invention is to provide an immunoassay through an antigen-antibody reaction for specifically and highly sensitively measuring PIVKA-II in serum or plasma by adding to reagents thrombin and/or an antibody reacting with human fibrin-like related substances.

Disclosure of the Invention:

To solve the problems described above, an immunoassay of PIVKA-II according to the present invention comprises the steps of adding to reagents thrombin and/or an antibody reacting with human fibrin-like related substances, and measuring PIVKA-II in serum or plasma.

The thrombin described above is preferably a thrombin-containing animal serum and/or purified thrombin, which may be heated or unheated.

In the present invention, the antibodies reacting with human fibrin-like related substances include, by way of example, anti-fibrinogen antibodies, anti-fibrin antibodies or the like, which are preferably polyclonal antibodies, especially those highly reacting with not only fibrinogen or fibrin but also fibrin-like related substances such as FDP, fibrinopeptide A or fibrinopeptide B. As the thrombin, there are used purified preparations of those derived from human beings or animals such as cows, pigs, sheep, horses, rabbits and chickens. Further, the use of a wide variety of thrombin-containing animal serums such as bovine serum, sheep

serum, porcine serum, horse serum, chicken serum and rabbit serum derived from animals different from species of animals immunized for obtaining the labeled antibodies or second antibodies may lead to reaction inhibition of anti-thrombin antibodies occurring as impurities in the labeled antibodies.

As the labeled antibodies or second antibodies used in the present invention, it is possible to use not only human polyclonal antibodies against prothrombin, F_1, F_2, or $F_1 + F_2$, but also human monoclonal antibodies against prothrombin, F_1, F_2, or $F_1 + F_2$. Here, F_1 and F_2 are peptides constituting prothrombin. Polyclonal or monoclonal antibodies prepared by immunization with synthetic peptides having the antigenecity of prothrombin can also be used.

The Examples of the present specification show application to electrochemiluminescence immunoassay. The present invention is also useful in trial of chemiluminescence assay, radioisotope assay or the like to achieve higher sensitivity. In the present invention, antibodies such as anti-fibrinogen antibody and anti-fibrin antibody reacting with fibrin-like related substances such as fibrinogen, fibrin, FDP, fibrinopeptide A and fibrinopeptide B are preferably obtained by immunization with human-derived fibrin-like related substances, but antibodies obtained by immunization with fibrin-like related substances such as animal-derived fibrinogen, fibrin or the like, and cross-reacting with human-derived fibrin-like related substances can also be used.

The antibodies such as anti-fibrinogen or anti-fibrin antibodies specific to fibrin-like related substances are preferably added to a reaction solution in the first reaction of the 2-step sandwich method. On the other hand, thrombin is preferably added to the labeled antibody solution or the second antibody solution in the second reaction, the adding amount thereof being preferably 1 to 50 NIH/ml. Those antibodies specific to fibrin-like related substances such as anti-fibrinogen or anti-fibrin antibodies, a purified thrombin, and an animal serum containing thrombin may be used singly or jointly as the occasion demands.

The animal serums containing thrombin, such as bovine serum, sheep serum, porcine serum, horse serum, chicken serum and rabbit serum derived from animals different from species of animals immunized for obtaining the labeled antibodies or second antibodies may be preferably added in an amount of 1 to 20 %. These animal serums derived from different species of animals may be blended, when necessary, with animal serums derived from the same animals as species of animals immunized for obtaining the labeled antibody or second antibody.

If the enzyme activity of thrombin is strong when adding thrombin to reagents, the immune reaction may be adversely affected, while if animal serums are added to those reagents containing the labeled antibodies or second antibodies, the stability of the reagents may be adversely affected, and therefore, a protease inhibitor for inhibiting the enzyme activity of thrombin

is preferably added to the reagents to which thrombin or animal serums are added.

As the protease inhibitor, it is possible to use inhibitors, mentioned on page 452 in "Rinsho Koso Handbook (Clinical Enzyme Handbook)" (1st ed., edited by Kitamura, Baba et al. and issued by Kodansha Scientific Co., on September 10, 1982), that is, plasma proteinous inhibitors, hirudine, benzamidine and synthetic inhibitor such as PMSF (phenylmethylsulfonyl fluoride), NPGB and or the like. However, these inhibitors are not sufficient for inhibiting the enzyme activity of thrombin, so it has been found that the enzyme activity is significantly reduced without losing its antigenecity even when a purified preparation of thrombin is subjected to heat treatment, e.g., at about 40 to 65 °C.

A commercial purified preparation of thrombin is to be stored primarily in a refrigerated or frozen form and not to be exposed to a high temperature. The heating temperature for thrombin used in the present invention is 30 to 70 °C, particularly preferably 40 to 60 °C, so that the heating time can be reduced to 15 to 60 minutes. As a matter of course, this heating is aimed at inactivating the enzyme activity of thrombin, and hence insofar as the enzyme can be inactivated without losing its antigenecity, the heating temperature and heating time are not limited to the above ranges.

If the animal serum derived from animals of species different from animals immunized for the labeled antibody is

previously heated for use as thrombin, the heating temperature is preferably 50 to 65 °C and the heating time is preferably 15 to 60 minutes. However, the heating time and heating temperature, needless to say, can be regulated without particular limitation in case of need. Further, the animal serum can be used without heating, if necessary.

Best Mode for Carrying out the Invention:

Hereinafter, the present invention is described by reference to the Examples, but these Examples are shown for illustrative purposes only and are not construed as restrictive.

Example 1 (an example of measurement by an electrochemiluminescence immunoassay in an automatic analyzer Picolumi 8220)

After 50 µl of a sample was added to 150 µl of a reaction solution, 25 µl of magnetic beads having anti-PIVKA-II monoclonal antibody immobilized thereon were added thereto. After they were reacted at 30 °C for 9 minutes, 350 µl of a Picolumi BF washing solution (10 mM Tris buffer) was added, and the magnetic beads trapped by a magnet were washed 3 times. To the magnetic beads thus subjected to the first reaction was added 200 µl of Ru-labeled antibody solution containing 1 µg/ml of a Ru-labeled anti-human prothrombin antibody (derived from rabbit), and these were allowed to react at 30 °C for 9 minutes. Likewise, the magnetic beads trapped by a magnet were washed 3 times with the

Picolumi BF washing solution. After addition of 300 μl of a Picolumi luminescent electrolytic solution containing 0.1 M tripropyl amine, the magnetic beads were sent to the surface of an electrode and the luminescence of Ru bound to the magnetic beads was measured, the amount of PIVKA-II in the sample being determined.

Reagent composition

Reaction solution: 50 mM Tris buffer (pH 7.8), 0.150 M NaCl, 0.01 % Tween 20, 0.1 % NaN_3, 5 % rabbit serum (heated).

Ru-labeled antibody solution: 50 mM Tris buffer (pH 7.8), 0.150 M NaCl, 0.01 % Tween 20, 0.1 % NaN_3, 1 mM PMSF, 1 μg/ml Ru-labeled anti-human prothrombin antibody (derived from rabbit), 5 % rabbit serum (heated).

(Preparation of solid-phase magnetic beads having anti-PIVKA-II monoclonal antibody)

1 ml of 30 mg/ml magnetic beads (4.5 microns) was put into a test tube and trapped with a magnet, and after the supernatant was discarded, 1 ml of 0.5 mg/ml anti-PIVKA-II monoclonal antibody (in 150 mM phosphate buffer, pH 7.8) was added to the magnetic beads, and these were allowed to react at room temperature for 1 day under stirring. After the magnetic beads were washed, 2 ml of 1 % BSA-Phosphate buffer was added thereto, and the magnetic beads were blocked for 1 day under stirring at room temperature. In case of use, the magnetic beads were diluted to 1 mg/ml with the 1 % BSA-Phosphate buffer.

(Preparation of Ru-labeled anti-human prothrombin antibody)

68 μl of Ru-complex compound of ruthenium-tri-dipyridyl modified with a succinimide group was added to 1 ml of 1 mg/ml anti-human prothrombin antibody immunized with rabbits, and these were allowed to react for 30 minutes under stirring at room temperature, and then the reaction was terminated by adding 50 μl of 2 M glycine, and further the sample was allowed to react for 10 minutes under stirring at room temperature. Finally, the sample was applied onto Sephadex G-25 (previously equilibrated with 10 mM phosphate buffer), and fractions of Ru-bound protein were collected. The Ru-labeled anti-human prothrombin antibody thus obtained was diluted to 1 μg/ml in case of use.

180 μg/ml of anti-human fibrinogen antibody (derived from rabbit) was added to each reaction solution, a control solution without the anti-human fibrinogen antibody (derived from rabbit) and 8 human serums being used to compare their specificity. Each serum was measured at n = 3, and the results are shown in Table 1. Those reagents with the anti-human fibrinogen antibody (derived from rabbit) show low dispersion in measured values and the absence of unspecific reaction.

Table 1

mAU/ml

	Control			C.V.	Addition of anti-fibrinogen antibody (180μg/ml)			C.V.
1	56	38	98	50.0%	26	23	24	6.3%
2	47	45	35	15.2%	27	27	28	2.1%
3	37	35	30	10.6%	26	33	29	12.0%
4	55	21	25	55.2%	20	22	19	7.5%
5	27	37	44	23.7%	15	19	15	14.1%
6	21	23	26	10.8%	19	19	20	3.0%
7	22	31	22	20.8%	21	20	18	7.8%
8	30	24	24	13.3%	24	24	24	0.0%

Example 2

In this example, the same reagent composition in Example 1 was used except that 10 NIH/ml of a purified preparation of bovine thrombin or human thrombin was added to the Ru-labeled antibody solution. This sample showing particularly highly unspecific reactions was selected and measured simultaneously for PIVKA-II at n = 10. The results of this sample and a control sample in which neither the bovine thrombin nor the human thrombin was added are shown in Table 2.

When the purified preparation of bovine or human thrombin was added to the reagent, the specificity of the sample was improved as compared with the control sample. This sample serum was centrifuged at 3000 rpm for 10 minutes and an obtained supernatant thereof showed 80 mAU/ml.

Example 3

In this example, the same reagent composition in

Example 1 was used except that 180 μg/ml anti-human fibrinogen antibody (derived from rabbit) was added to the reaction solution and 10 NIH/ml bovine thrombin was added to the Ru-labeled antibody solution. This sample showing highly unspecific reaction was measured simultaneously for PIVKA-II at n = 10. The results of this sample and a control sample to which neither the anti-human fibrinogen antibody (derived from rabbit) nor the bovine thrombin was added are shown in Table 2 with the results in Example 2. Addition of both the anti-human fibrinogen antibody (derived from rabbit) and bovine thrombin leads to an increase in specificity of PIVKA-II much more in comparison with addition of bovine thrombin alone.

Table 2

mAU/ml

	Control	Addition of human thrombin	Addition of bovine thrombin	Use of both bovine thrombin and anti-human fibrinogen antibody
1	365	121	99	97
2	219	83	145	80
3	158	83	112	79
4	209	88	107	89
5	202	95	119	72
6	150	154	104	79
7	247	84	110	80
8	133	110	103	82
9	166	121	94	85
10	245	92	103	100
Mean	209.4	103.1	109.6	84.3

Example 4

500 NIH of a purified preparation of bovine thrombin was added to 1 ml of 50 mM Tris buffer (0.15 M NaCl, pH 7.8) and heated at 50 °C for 30 minutes in a thermostatic water bath. In this example, the same reagent composition in Example 1 was used except that 180 μg/ml anti-human fibrinogen antibody (derived from rabbit) was added to the reaction solution and the heat-treated purified preparation of bovine thrombin was added at a concentration of 5 NIH/ml to the Ru-labeled antibody solution. Using this sample with a high unspecific reaction, PIVKA-II was measured. The results of this sample and a control sample to which neither the anti-human fibrinogen antibody (derived from rabbit) nor the heated bovine thrombin was added are shown in Table 3. As is evident from the results in Table 3, the inhibitory effect of the sample solutions in this example against the unspecific reaction was exhibited similarly in Example 3. The enzyme activity of this heated thrombin, as measured using Chromozyme TH (Boehringer), was reduced to 1/5 compared with that of the unheated thrombin.

Table 3

mAU/ml

	Control	Use of both heated bovine thrombin and anti-human fibrinogen antibody
1	133	105
2	154	87
3	152	92
4	219	132
5	150	52
6	100	87
7	137	98
8	125	90
9	162	89
10	127	86
Mean	145.9	91.8

Examples 5 and 6

In this example, the same reagent composition in Example 1 was used except that 5 % unheated rabbit serum (a control where the final concentration of the rabbit serum was 10 %), 5 % unheated horse serum (Example 5), or 5 % unheated sheep serum (Example 6) was added to Ru-labeled antibody solution, respectively. This sample showing highly unspecific reaction was measured for its inhibitory effect against the unspecific reaction, and the results are shown in Table 4.

Table 4

mAU/ml

	Control	Rabbit serum + horse serum	Rabbit serum + sheep serum
1	208	114	114
2	190	104	109
3	196	107	120
4	179	135	136
5	242	170	132
Mean	203	126	122

As compared with the control, the unspecific reaction was inhibited in the sample to which the horse serum or sheep serum was added. This sample showing highly unspecific reaction was centrifuged at 3000 rpm for 10 minutes and an obtained supernatant thereof showed 74 mAU/ml.

Capability of Exploitation in Industry:

As described above, according to the present invention, PIVKA-II in serum or plasma can be measured specifically and highly sensitively by adding to reagents thrombin and/or an antibody reacting with human fibrin-like related substances.

CLAIMS

1. An immunoassay of PIVKA-II in serum or plasma, comprising the steps of:

adding to reagents thrombin and/or an antibody reacting with human fibrin-like related substances, and

measuring PIVKA-II in serum or plasma.

2. The immunoassay according to claim 1, wherein a thrombin-containing animal serum and/or purified thrombin are used as the thrombin.

3. The immunoassay according to claim 1 or 2, wherein an anti-fibrinogen antibody and/or an anti-fibrin antibody are used as the antibody reacting with human fibrin-like related substances.

FOR OFFICIAL USE ONLY

ABSTRACT

The present invention provides an immunoassay for specifically and highly sensitively measuring PIVKA-II in serum or plasma through antigen-antibody reaction by adding to reagents an animal serum containing thrombin and/or an antibody reacting with human fibrin-like related substances. The immunoassay of the invention comprises the steps of adding to reagents thrombin and/or an antibody reacting with human fibrin-like related substances, and measuring PIVKA-II in serum or plasma.

DECLARATION AND POWER OF ATTORNEY FOR U.S. PATENT APPLICATION

( ) Original ( ) Supplemental ( ) Substitute (X) PCT ( ) Design

As a below named inventor, I hereby declare that: my residence, post office address and citizenship are as stated below next to my name; that I verily believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Title: IMMUNOASSAY OF PIVKA-II

of which is described and claimed in:
( ) the attached specification, or
( ) the specification in the application Serial No. _____ filed _____;
and with amendments through _____ (if applicable), or
(X) the specification in International Application No. PCT/ JP00/03550, filed on June 1, 2000, and as amended on _____ (if applicable).

I hereby state that I have reviewed and understand the content of the above-identified specification, including the claims, as amended by any amendment(s) referred to above.

I acknowledge my duty to disclose to the Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I hereby claim priority benefits under Title 35, United States Code, §119 (and §172 if this application is for a Design) of any application(s) for patent or inventor's certificate listed below and have also identified below any application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

COUNTRY	APPLICATION NO.	DATE OF FILING	PRIORITY CLAIMED
JAPAN	11-354862	14/12/1999	YES

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose information material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application.

APPLICATION SERIAL NO.	U.S. FILING DATE	STATUS: PATENTED, PENDING, ABANDONED

ATTACHMENT B

(6)

And I hereby appoint Michael R. Davis, Reg. No. 25,134; Matthew M. Jacob, Reg. No. 25,154; Jeffrey Nolton, Reg. No. 25,408; Warren M. Cheek, Jr., Reg. No. 33,367; Nils E. Pedersen, Reg. No. 33,145 and Charles R. Watts, Reg. No. 33,142, who together constitute the firm of WENDEROTH, LIND & PONACK, L.L.P., attorneys to prosecute this application and to transact all business in the U.S. Patent and Trademark Office connected therewith.

I hereby authorize the U.S. attorneys named herein to accept and follow instructions from ______________________________ ______________________________ as to any action to be taken in the U.S. Patent and Trademark Office regarding this application without direct communication between the U.S. attorneys and myself. In the event of a change in the persons from whom instructions may be taken, the U.S. attorneys named herein will be so notified by me.

Send Correspondence to

WENDEROTH, LIND & PONACK, L.L.P.
2033 K Street, N.W., Suite 800
Washington, DC 20006

Direct Telephone Calls to:

WENDEROTH, LIND & PONACK, L.L.P.
Area Code (202) 721-8200

Direct Facsimile Messages to:
Area Code (202) 721-8250

1-00

Full Name of First Inventor	FAMILY NAME KANASHIMA	FIRST GIVEN NAME Motohito	SECOND GIVEN NAME	
Residence & Citizenship	CITY Toride-shi	STATE OR COUNTRY Ibaraki	COUNTRY OF CITIZENSHIP Japan JPX	
Post Office Address	ADDRESS 10-30-307, Toride 2-chome,	CITY Toride-shi	STATE OR COUNTRY Ibaraki	ZIP CODE 302-0004

2-00

Full Name of Second Inventor	FAMILY NAME ASAI	FIRST GIVEN NAME Tomohide	SECOND GIVEN NAME	
Residence & Citizenship	CITY Ushiku-shi	STATE OR COUNTRY Ibaraki	COUNTRY OF CITIZENSHIP Japan JPX	
Post Office Address	ADDRESS 31-10, Minami 7-chome	CITY Ushiku-shi	STATE OR COUNTRY Ibaraki	ZIP CODE 300-1222

3-00

Full Name of Third Inventor	FAMILY NAME TAKAHASHI	FIRST GIVEN NAME Hiroki	SECOND GIVEN NAME	
Residence & Citizenship	CITY Ikoma-shi	STATE OR COUNTRY Nara	COUNTRY OF CITIZENSHIP Japan JPX	
Post Office Address	ADDRESS 207-83, Higashi-Ikoma 2-chome	CITY Ikoma-shi	STATE OR COUNTRY Nara	ZIP CODE 630-0213

4-00

Full Name of Fourth Inventor	FAMILY NAME ASAI	FIRST GIVEN NAME Yoshiyuki	SECOND GIVEN NAME	
Residence & Citizenship	CITY Yono-shi	STATE OR COUNTRY Saitama	COUNTRY OF CITIZENSHIP Japan JPX	
Post Office Address	ADDRESS 10-2, Kamiochiai 1-chome	CITY Yono-shi -	STATE OR COUNTRY Saitama	ZIP CODE 338-0001

Full Name of Fifth Inventor	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME	
Residence & Citizenship	CITY	STATE OR COUNTRY	COUNTRY OF CITIZENSHIP	
Post Office Address	ADDRESS	CITY	STATE OR COUNTRY	ZIP CODE
Full Name of Sixth Inventor	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME	
Residence & Citizenship	CITY	STATE OR COUNTRY	COUNTRY OF CITIZENSHIP	
Post Office Address	ADDRESS	CITY	STATE OR COUNTRY	ZIP CODE
Full Name of Seventh Inventor	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME	
Residence & Citizenship	CITY	STATE OR COUNTRY	COUNTRY OF CITIZENSHIP	
Post Office Address	ADDRESS	CITY	STATE OR COUNTRY	ZIP CODE

I further declare that all statements made herein of my own knowledge are true, and that all statements on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

1st Inventor Motohito Kanashima Date April 11, 2001

2nd Inventor Tomohide Asai Date April 11, 2001

3rd Inventor Hiroki Takahashi Date April 20, 2001

4th Inventor Toshiyuki Date April 10, 2001

5th Inventor _____ Date _____

6th Inventor _____ Date _____

7th Inventor _____ Date _____

The above application may be more particularly identified as follows:

U.S. Application Serial No. _____ Filing Date _____

Applicant Reference Number _____ Atty Docket No. _____

Title of Invention _____

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